

Evaluation Of the Effectiveness of Non-Steroidal Anti-Inflammatory Drugs (NSAIDs) According to Their Cyclooxygenase Selectivity in Neuropathic Pain Model in Rats (Experimental Study)

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Abstract

Aim: Recent studies on neuropathic pain treatment and mechanisms reveal that NSAIDs, which are mostly used in types of pain related to inflammation, may also be effective in neuropathic pain. In this experimental study, we created a neuropathic pain model in rats with sciatic nerve ligation. We investigated the effectiveness of lornoxicam (nonselective COX enzyme inhibitor), meloxicam (selective COX-2 enzyme inhibitor), and dexametopfen (nonselective COX enzyme inhibitor) on mechanical hyperalgesia, which is a clinical finding of neuropathic pain and can be measured objectively and numerically with electronic von Frey (evF) according to their cyclooxygenase enzyme (COX) selectivity. Thus, we aimed to contribute to the literature and clinical practice by demonstrating the effectiveness of COX selectivity in the treatment of neuropathic pain.

Materials and Methods: 35 male Wistar-albino rats with normal motor activity were included in the study. The animals were randomly divided into 5 groups as control (Group I n=7), lornoxicam (Group II n=7), meloxicam (Group III n=7), dexametopfen (Group IV), and sham surgery (Group V). The Chronic Constriction Injury (CCI) - induced neuropathy model described by Bennett and Xie was applied. As described in this model, after the sciatic nerve of the right hind legs of the rats was surgically exposed, three loose knots were tied with 4/0 catgut at 1 mm intervals from 4 different places. In the group that underwent sham surgery, the sciatic nerve was re-closed without any procedure after being exposed. In this way, lornoxicam 1.3 mg/kg, meloxicam 5.8 mg/kg, and dexametopfen 15 mg/kg were administered intraperitoneally to the groups that developed neuropathic pain. Drug applications were performed in a single-blind manner. Mechanical hyperalgesia measurements were made with an EVF device and recorded. The study was completed without any problems with 35 subjects.

Results: In rats with neuropathic pain, a significant difference was found between the control group that received physiological serum and the groups that received COX inhibitors at all measurement times ($p < 0.008$). When the drug-administered groups were compared in pairs; Intraperitoneal dexametopfen administration was found to be statistically more significant than intraperitoneal lornoxicam or meloxicam administration at 30th minute ($p < 0.008$). Intraperitoneal lornoxicam administration was found to be statistically more significant than intraperitoneal meloxicam or dexametopfen administration at 150th and 180th minute ($p < 0.008$).

Conclusion: COX inhibitors are effective in preventing mechanical hyperalgesia in neuropathic pain.

Keywords: lornoxicam; meloxicam; dexametopfen; neuropathic pain models in rats; neuropathic pain

Introduction

Neuropathic pain is defined as “pain resulting from a disease or lesion affecting the somatosensory system” (Treede RD, 2008). It is a chronic condition when not treated promptly and correctly. A better understanding of the mechanisms of neuropathic pain, which is completely different from other types of pain, has made it easier for us to approach this group of diseases that are difficult to diagnose and treat. Antidepressants, anticonvulsants, opioids, local anesthetics and capsaicin, local preparations and interventional methods are used in the treatment of neuropathic pain. Simple analgesics, such as paracetamol and nonsteroidal anti-inflammatory drugs (NSAIDs), are generally ineffective in the treatment of neuropathic pain (Namaka M. 2004, Namaka M. 2009). However, in experimental studies conducted in recent years, the effectiveness of simple analgesics in neuropathic pain models has been evaluated, and it has been argued that paracetamol and NSAIDs will increase the effectiveness of these drugs, especially when added to other drugs used in the treatment of neuropathic pain (McCormack KJ. 1994, Eroğlu L 2002). The antihyperalgesic effect of paracetamol increases dose-dependently and shows a synergistic effect when used together with other drugs used in the treatment of neuropathic pain (Raffa R. 2010, Dani M. 2007, Bonnefont J. 2003). Experimental studies have shown that NSAIDs reduce mechanical hyperalgesia and their effectiveness varies according to their COX selectivity (Kimura S. 2009). Meloxicam, a selective COX-2 inhibitor, was found to be more effective than ibuprofen, a selective COX-1 inhibitor, in rats with diabetic neuropathy. Lornoxicam, a nonselective COX inhibitor, was reported to be more effective than proxicam in the same group and to reduce mechanical hyperalgesia (Bianchi M. 2002). We did not come across a study in the literature evaluating the effectiveness of dexketoprofen, which has been widely used in our country in recent years, on mechanical hyperalgesia. In this study, lornoxicam, meloxicam and dexketoprofen were used in rats

in an experimental neuropathic pain model. We aimed to . evaluate the effects of profen on mechanical hyperalgesia.

Material And Methods

This study was conducted in SUDAM laboratories and Selcuk University Selcuklu Medical Faculty Pharmacology Department laboratory after receiving approval from Selcuk University Meram Medical Faculty Experimental Animal Research Center (SUDAM) ethics committee with the number 2011-050.

Subjects:

35 Wistar-albino male rats were included in the study. The rats weighed between 315-520 gr and had normal motor activity. The floor of the cages where the experimental animals were housed was kept soft with sawdust in order to minimize possible painful mechanical stimulation. Rats were placed in cages in numbers of maximum 4. The rooms where the cages were located were standardized to prevent external light from passing through and to have a 12-hour light and 12-hour dark cycle, an ambient temperature of $22 \pm 2^\circ\text{C}$ and a humidity of 70-75%. They were fed with standard rat food and tap water.

Preparation of the subjects:

Before the surgical procedure, the responses of all animals to mechanical stimulation were measured with evF to establish baseline values and recorded. All rats, whose feeding was stopped 12 hours before the operation and who were allowed to drink only water, were anesthetized with a mixture of 50 mg/kg intraperitoneal ketamine 10% (Ketalar®) and 10mg/kg intraperitoneal xylazine 2% (Rompun®) (Figure 3.1).



Figure 3.1: Administration of anesthesia with an intraperitoneal ketamine-xylazine mixture.

After anesthesia induction, the CCI-related neuropathy model described by Bennett and Xie was applied. For this purpose, the hair covering the right thigh region of the rats was shaved and cleaned, and the opened area was wiped with povidone iodine. The skin was cut with a scalpel so that

it would extend parallel to the thigh. The sciatic nerve was reached with blunt dissection along the M. Biceps femoris. Three loose knots were tied with 4/0 catgut at 1 mm intervals from 4 different places on the exposed sciatic nerve (Figure 3.2).



Figure 3.2: Ligation of the sciatic nerve.

The incision was closed in layers with 3/0 silk (Figure 3.3).



Figure 3.3: Closure of the skin.

The rats were waited for 21 days for neuropathic pain to develop. In the sham (pseudo-surgery) group, similar surgery was applied without affecting the sciatic nerve, and the skin was closed after the sciatic nerve was exposed.

Experimental groups:

Animals that developed neuropathic pain were randomly divided into control, lornoxicam, meloxicam, and dexketoprofen groups (Table 3.1). Drug administrations were performed in a single-blind manner.

Groups	Explanation	Number of subjects	Applied content
1	Control group	7	2 ml saline
2	Lornoxicam group	7	1.3 mg/kg lornoxicam
3	Meloxicam group	7	5.8 mg/kg Meloxicam
4	Dexketoprofen group	7	15 mg/kg deksketoprofen
5	Sham group	7	none

Table 3.1: Working groups and drugs administered

Preparation And Administration Of Drugs

Xefo vial (Lornoxicam, Abdi İbrahim, 4 mg/ml), Meloks ampoule (Meloxicam, Nobel İlaç Sanayi, 10 mg/ml) and Arvels ampoule (Dexketoprofen, İbrahim Etem Ulagay, 25 mg/ml) were used as drugs. The drugs were drawn into 5 ml syringes in doses appropriate to the weights of the rats. They were completed to 10 ml volume with physiological serum. The prepared drugs were injected intraperitoneally with a 22 G needle into the right lower quadrants of the rats that were held with the appropriate method.

Measurement With Evf

After the surgical intervention and waiting 21 days for the development of neuropathic pain, the weights of the rats were measured again before the drug injection. In the extremity where the CZ-induced neuropathy model was applied, mechanical hyperalgesia was measured with the evF device at minute 0 (before drug or physiological serum application) and at minutes 30, 60, 90, 120, 150 and 180 after the application of the drugs. After the device was calibrated, each rat was placed in open-topped, transparent-sided cages with holes suitable for mechanical stimulation to the plantar surface of the foot. Measurements were started on rats that got used to the environment and exhibited natural behaviors. All

measurements were performed by the same researcher in order to avoid hand changes. The evF fiber was contacted at a right angle to the mid-

plantar surface of the right hind extremity of the rats with the help of the mirror under the measurement cages (Figure 3.4).



Figure 3.4: Measurement with electronic von Frey

The applied pressure force was gradually increased. The increase in the applied force was observed graphically and numerically on the computer screen (Figure 3.5).

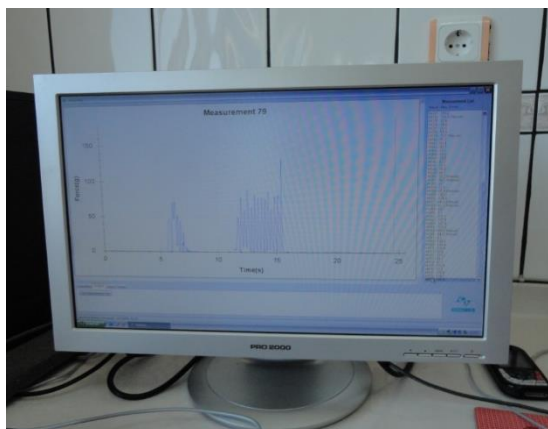


Figure 3.5: The increase in applied force Graphical and numerical detection

The numerical value when the rat withdrew its paw was recorded by viewing it on the computer screen. The measurements were repeated and the values were confirmed. The withdrawal made by the animal during spontaneous movement, independent of pain, was evaluated as false positive and was not considered significant. The cut-off value was determined as 200 g since it would damage the rat's paw and could also falsely affect the reliability of the measurement. After the measurements, the rats were sacrificed under anesthesia by the cervical dislocation method.

Statistical Analysis

After calculating the % maximal possible effects (MPE) of the withdrawal threshold values measured at the end of the study, the new values obtained were used for statistical analysis. The following formula and the 200 g cut-off value were used for this calculation. Statistical analysis was performed using the SPSS v16.0 (SPSS Inc., Chicago, IL, USA) package

program. Continuous variables were shown as mean±standard deviation. Categorical variables were expressed as frequency percent. It was given as a series. The difference between all groups was made with Kruskal-Wallis one-way variance analysis. A value smaller than $P < 0.05$ was considered significant. In the minutes when it was significant, the groups were compared in pairs. Bonferroni correction was applied for this comparison and Mann-Whitney U test was used. Bonferroni correction coefficient was taken as 6. Therefore, a value smaller than $P < 0.008$ was considered statistically significant. Preoperative and postoperative weights were investigated with Wilcoxon test. A P value smaller than 0.05 was considered statistically significant.

Results

There was no significant difference between the weights measured before and after surgery of Wistar Albino male rats included in the study ($p < 0.05$) (Table 4.1). No injury or autotomy was observed.

WEIGHT (gr)	Group I	Group II	Group III	Group IV	Group V	P
Weight before surgery	371,85±42,95	354,85±35,41	370,85±43,75	378,28±78,49	372,28±40,48	1,00
Weight after surgery	372,28±42,80	354,42±37,72	372,00±45,10	378,85±77,88	372,57±42,01	1,00

Table 4.1: Rat weights.

Effectiveness of the CCI model:

The measurement values in the control group (Group I) were numerically lower than those in the sham group (Group V). When the measurement

values of Group I and Group V were compared statistically to evaluate the effectiveness of the CCI model, a significant difference was found ($p < 0.008$) (Table 4.2). These data showed that neuropathic pain developed in rats that received CCI and that the surgery was effective.

Time (min.)	Group I (n=7)	Group V (n=7)	P
30. min.	1,09±0,85	4,62±1,65	0,002*
60. min.	1,12±1,06	4,59±2,20	0,003*
90. min.	0,80±0,44	4,67±2,07	0,002*
120. min.	0,78±0,36	5,01±2,16	0,002*
150. min.	1,34±0,81	5,79±1,46	0,002*
180. min.	1,061±0,55	6,21±2,08	0,002*

* $p < 0.008$ Significant difference

Table 4.3: Measurement values of all groups (Mean± SD).

Effectiveness of drugs compared in the study:

Measurement values after drug applications are shown in (Table 4.3, Graph 4.1).

When Group II, which received lornoxicam, was examined; Measurement values were higher at all evaluation times compared to Group I ($p < 0.008$) (Table 4.4).

Effect of Lornoxicam:

Time (min.)	Group I (n=7)	Group II (n=7)	P
30. min.	1,09±0,85	10,22±4,75	0,002*
60. min.	1,12±1,06	24,58±3,34	0,002*
90. min.	0,80±0,44	22,94±7,32	0,002*
120. min.	0,78±0,36	33,36±4,81	0,002*
150. min.	1,34±0,81	50,86±8,68	0,002*
180. min.	1,061±0,55	71,51±1,90	0,002*

* $P < 0.008$ Significant difference

Table 4.4: Group I and Group II measurement values (Mean± SD).

When Group II measurement values were compared with Group III, it was observed that they were more effective at 150 and 180 minutes. When compared with Group IV, it was observed that Group IV was more

effective at 30 minutes, while Group IV was more effective at 150 and 180 minutes ($p < 0.008$) (Table 4.5, Table 4.6).

Time (dk)	Group II (n=7)	Group III (n=7)	P
30. min.	10,22±4,75	10,74±5,31	0,848
60. min.	24,58±3,34	30,29±8,89	0,180
90. min.	22,94±7,32	27,09±9,51	0,277
120. min.	33,36±4,81	28,31±1,12	0,406
150. min.	50,86±8,68	23,81±1,39	0,003*
180. min.	71,51±1,90	33,05±1,21	0,004*

* $P < 0.008$ Significant difference

Table 4.5: Group II and Group III measurement values (Mean± SD).

Time (min.)	Group II (n=7)	Group IV (n=7)	P
30. min.	10,22±4,75	25,22±5,05	0,003*
60. min.	24,58±3,34	16,15±7,73	0,025
90. min.	22,94±7,32	21,91±3,08	0,277
120. min.	33,36±4,81	25,62±8,2	0,018
150. min.	50,86±8,68	22,20±4,00	0,002*
180. min.	71,51±1,90	27,84±6,65	0,002*

* $P < 0.008$ Significant difference

Table 4.6: Group II and Group IV measured values (Min.± SD).

Effect of Meloxicam:

When Group III, where meloxicam was applied, was examined; the measurement values were higher at all evaluation times compared to Group I ($p<0.008$) (Table 4.7).

Time (min.)	Group I (n=7)	Group III (n=7)	P
30. min.	1,09±0,85	10,74±5,31	0,002*
60. min.	1,12±1,06	30,29±8,89	0,002*
90. min.	0,80±0,44	27,09±9,51	0,002*
120. min.	0,78±0,36	28,31±1,12	0,002*
150. min.	1,34±0,81	23,81±1,39	0,002*
180. min.	1,061±0,55	33,05±1,21	0,002*

* P <0,008 Significant difference

Table 4.7: Group I and Group III measurement values (Min.± SD).

When compared to Group IV, it was observed that Group IV was more effective at the 30th minute ($p<0.008$) (Table 4.8).

Time (dk)	Group III (n=7)	Group IV (n=7)	P
30. min.	10,74±5,31	25,22±5,05	0,004*
60. min.	30,29±8,89	16,15±7,73	0,018
90. min.	27,09±9,51	21,91±3,08	0,142
120. min.	28,31±1,12	25,62±8,2	0,949
150. min.	23,81±1,39	22,20±4,00	0,749
180. min.	33,05±1,21	27,84±6,65	0,565

Table 4.8: Group III and Group IV measured values (Mean± SD).

Effect of Dextetoprofen:

When Group IV, where dextetoprofen was applied, was examined; the measurement values were higher at all evaluation times compared to Group I ($p<0.008$) (Table 4.9) (Figure 4.1.)

Time (dk)	Group I (n=7)	Group IV (n=7)	P
30. min.	1,09±0,85	25,22±5,05	0,002*
60. min.	1,12±1,06	16,15±7,73	0,003*
90. min.	0,80±0,44	21,91±3,08	0,002*
120. min.	0,78±0,36	25,62±8,2	0,002*
150. min.	1,34±0,81	22,20±4,00	0,002*
180. min.	1,061±0,55	27,84±6,65	0,002*

*P<0,008 Significant difference

Table 4.9: Group I and Group IV measured values (Mean± SD).

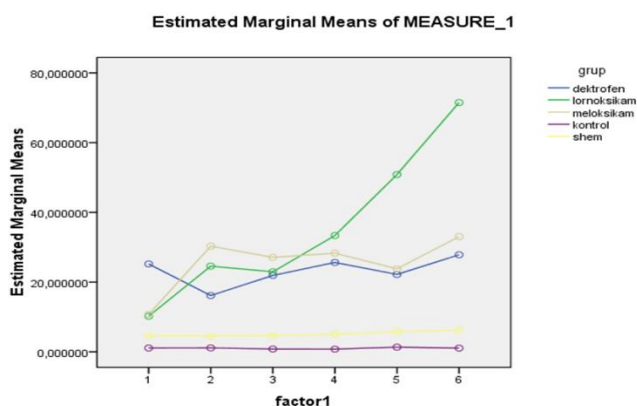


Figure 4.1: Metric values of all groups.

Discussion

The significant differences in the regions where it originates and its symptoms, inadequacies in diagnosis, the lack of a complete understanding of the mechanism, and the disregard of factors that increase pain such as depression and anxiety are among the most important reasons why neuropathic pain treatment is difficult (Tuncer A. 2003, Attal N. 2005). Despite the better understanding of its mechanisms and the diversification of drug groups used in its treatment with experimental and clinical studies conducted in recent years, neuropathic pain is still a difficult pain to treat. Today, the main limiting factor in the treatment of neuropathic pain is that frequently used drugs such as anti-depressants and anti-convulsants are off-label (use outside the approved area of use of a drug). Beneficial effects have been shown for agents such as opioids, tramadol, topical medications (lidocaine, capsaicin) used in treatment, and mexiletine, baclofen, ketamine and NSAIDs have been used in some treatments. Despite the use of all these agents, significant pain reduction in patients with neuropathic pain is less than half (Eisenberg E. 2005). Various algorithms and guidelines have been published on the treatment of neuropathic pain.

Namaka M. et al.'s 2009 report on neuropathic pain management outlined four phases based on scanning all articles and researchers' experiences. Antidepressants, antiepileptics, and topical analgesics are advised first. The second phase should utilise narcotic analgesics and refractory medicines, and the third step should use combination therapy. Surgical procedures are indicated as the fourth step if these medications fail to relieve discomfort. Adjuvant analgesic therapy with NSAIDs is possible at all stages (Namaka M. 2009). Despite emerging treatments, no single treatment works for all neuropathic pain. Chronic neuropathic pain is rarely relieved by monotherapy, despite the goal of treating it with one drug. NSAIDs may cure neuropathic pain by modulating several central and peripheral components of pain processes (Namaka M. 2009, Bianchi M. 2002). COX selectivity was used to assess NSAID efficacy in rats with experimental neuropathic pain. Experimental models for animal neuropathic pain use peripheral nerve mechanical damage. Chronic constriction injury (CCI), partial tight ligation (PSL), and spinal nerve ligation (SSL) are the most common partial denervation models, along with the streptozocin-induced diabetic neuropathy model (Ulugöl A. 2012). The CCC model by Bennett and Xie involves tying knots around the sciatic nerve to cause chronic constriction injury and inhibit superficial epineurial vascularization (Bennett GJ. 1988). Four ligatures at 1 mm intervals strangle the sciatic nerve by causing intraneuronal oedema. Compression disrupts nerve axons. Destruction of neural structure distal to compression is typical. Due to spontaneous discomfort, rats exhibit several behaviours. These include mild-moderate autotomy (self-attack on the lesioned leg resulting in amputation), protection, excessive licking, limping, and not stepping on it. Autotomy rats are not used in experiments. Detect cold allodynia and hyperalgesia from noxious thermal and mechanical stimulation. Unilateral symptoms. Allodynia, hyperalgesia, and spontaneous pain-related behavioural markers peak two weeks after surgery and persist 2-3 months. We used the CCI neuropathic pain model on rats in this work. Three weeks passed before we measured, including the second week following surgery when neuropathic pain peaked. We compared the sham group, which underwent surgery without harming the sciatic nerve, to the control group to determine if our surgery was effective. Control group (Group I) measurement values were lower than sham group (Group V). Statistics showed a considerable disparity between Group I and Group V measurement values. ($p < 0.008$) (Table 4.2). All of these results revealed that CPR caused neuropathic pain in rats and that our operation worked. There were no autotomy attacks on the limb with the nerve lesion. This helped apply CPR correctly. Rats and

mice, the most common pain study rodents after humans, are challenging to assess for pain threshold and analgesia. In identical pain situations, nonverbal subjects often demonstrate simple reflexes or motor behaviours like escape. Rats exhibit behavioural indications of neuropathic pain such as allodynia and hyperalgesia, as evidenced by numerous studies. The evF approach is the most recommended for assessing mechanical hyperalgesia in neuropathic pain in experimental research. Classical von Frey flame measurements are negatively influenced by external conditions like heat and humidity, which change flame properties. Therefore, an electronic von Frey device is recommended for measurements (Möller KA. 1998). Similar to previous investigations, we used an electronic von Frey device to quantify functional pain and make measurements. We used drug pharmacokinetics to determine measurement periods following administration. Dexketoprofen trometamol reaches C-max in 20 minutes after IM and 30 minutes after oral dosing. Distribution half-life is 0.35 hours and elimination half-life is 1-2.7 hours (Barbanoj MJ. 2001). Complete absorption of meloxicam after IM injection. Plasma concentrations depend on dosage. Plasma levels stabilise in 3-5 days. Within 60 minutes of 15 mg IM injection, C-max is 1.62 mg/L (Martindale 2007, Megan S. K. 2006). Lornoxicam Cmax is 25 minutes after IM injection. Given this, we measured at 30, 60, 90, 120, 150, and 180 minutes following drug delivery. In our 30th minute tests, dexketoprofen achieved C-max the fastest and was statistically more significant and effective than the other two medications. NSAIDs block PG and leukotrienes to relieve pain. Recently, NSAIDs have been shown to affect the central nervous system. Different CNS areas are affected by NSAIDs. In animals, they diminish hyperalgesia generated by esi and NMDA actions at the spinal level. NSAIDs can pass the blood-brain barrier and block PG production in opioid-related noradrenergic pathways that inhibit CNS pain (Steiner AA. 2001). In animal models, NSAIDs affect the hypothalamus, thalamus, and periaqueductal grey matter (Katz JA. 2000). The fact that diclofenac's analgesic effect can be reversed with naloxone and that it reduces heroin addicts' withdrawal symptoms suggests that NSAIDs work through central opioid pathways. The doses of lornoxicam 1.3 mg/kg, meloxicam 5.8 mg/kg, and dexketoprofen 15 mg/kg that are most commonly used in pain studies and can reach the central nervous system were chosen. A rat study found that the spinal COX enzyme supports peripheral analgesia via the central pathway. NSAIDs can exert central effects via opioidergic, serotonergic, NMDA, and excitatory amino acids (McCormack K. 1994, Eroğlu L. 2002). Dirig MD. Et al. found that COX inhibitors can be given systemically and spinally to block the initial pain component in thermal hyperalgesia caused by tissue injury, but they must be given systemically to treat established thermal hyperalgesia. In hyperalgesia, systemic COX inhibitors worked similarly.

In their investigation on mice with diabetic neuropathic pain, Kimura S. et al. compared 30 mg/kg ibuprofen to 3, 10, and 30 mg/kg meloxicam. They found that dosages above 3 mg/kg significantly enhanced paw withdrawal threshold. Ibuprofen was minor (Kimura S. 2009). Like our trial, meloxicam was beneficial in neuropathic pain. Meloxicam in combination reduces neuropathic pain, according to research. It reduced neuropathic pain better when administered with aminoguanidine hydrochloride than alone, according to Dudhgaonkar SP. et al. (Dudhgaonkar SP 2007). No meloxicam combinations were used in our study.

Similar to our investigation, Takahashi M. et al. found that systemic meloxicam significantly improved tactile allodynia in mice following L5 spinal nerve injury compared to vehicle treatment (Takahashi M. 2005). Lornoxicam demonstrated great tolerability and a higher analgesic efficacy than placebo in a double-blind multicentric parallel group study

of 171 individuals with acute sciatic or lumbosciatic pain (Herrmann WA. 2009). Lornoxicam had similar effects to diclofenac in this study. Hu Y. et al. compared lornoxicam to amitriptyline, a common neuropathic pain therapy, on 60 male rats after L5 spinal incision and sham surgery. Amitriptyline treated mechanical allodynia, depression-related behaviours, and cognitive functioning, while lornoxicam only treated mechanical allodynia (Hu Y. 2010). It solely affected mechanical allodynia in our investigation. Lornoxicam, piroxicam, and meloxicam were tested for pain in rats with 10% formaldehyde-damaged tails by Bianchi et al. These medicines greatly reduced hyperalgesia but did not modify thermal pain summation. However, lornoxicam alone prevented hyperalgesia (Bianchi M 2002). As in this study, lornoxicam outperformed meloxicam at 150 and 180 minutes. In rats and mice, Cabra F. and colleagues found dexketoprofen to be strong anti-inflammatory, analgesic, and antipyretic (Cabr  F. 1998). No study on dexketoprofen and neuropathic pain was found. However, a study found that dexketoprofen synergistically improves pain therapy when combined with other drugs. Miranda HF. and colleagues examined acute tonic, phasic, and inflammatory pain in mice with dexketoprofen, morphine, and paracetamol. Dexketoprofen synergised with both medicines in all three experiments (Miranda HF. 2007). Like Miranda HF and colleagues, The combination with tramadol was found to be synergistic as antinociceptive and antiexudative (Miranda HF. 2012).

Conclusion

In conclusion, our research indicates that COX inhibitors, such as lornoxicam (1.3 mg/kg), meloxicam (5.8 mg/kg), and dexketoprofen (15 mg/kg), have substantial antihyperalgesic effects in rats suffering from neuropathic pain caused by chronic constriction injury (CCI). Of these, dexketoprofen exhibited the swiftest onset of action, being most efficacious at 30 minutes, although lornoxicam displayed greater efficacy at 150 and 180 minutes in comparison to meloxicam and dexketoprofen. Due to the insufficient evidence on dexketoprofen in neuropathic pain models, additional research is required to investigate its complete potential. This study, focussing on single-dose administration, necessitates further long-term investigations to validate the enduring efficacy of these COX inhibitors in the management of neuropathic pain.

References

1. Attal N, Cruccu G, Baron R. (2005). EFNS guidelines on the pharmacological treatment: an evidence based proposal. *Pain*. 118:289-305.
2. Barbanoj MJ, Antonijoan RM, Gich I.(2001). Clinical pharmacokinetics of dexketoprofen. *Clin Pharmacokinet*. 40:245-262.
3. Bennett GJ, Xie YK. (1988). A peripheral mononeuropathy in rat that produces disorders of pain sensation like those seen in man. *Pain*. 33:87-107.
4. Bianchi M, Panerai AE. (2002). Effects of lornoxicam, piroxicam, and meloxicam in a model of thermal hindpaw hyperalgesia induced by formalin injection in rat tail. *Pharmacol Res*. 45(2):101-105.
5. Bonnefont J, Alloui A, Chapuy E. (2003). Orally administered paracetamol does not act locally in the rat formalin test: evidence for a supraspinal, serotonin-dependent antinociceptive mechanism. *Anesthesiology*. 99:976-981.
6. Cabr  F, Fern ndez MF, Calvo L, Ferrer X, Garc a ML. et al. (1998). Analgesic, antiinflammatory, and antipyretic effects of S(+)-ketoprofen in vivo. *J Clin Pharmacol*. 38(12 Suppl):3S
7. Dudhgaonkar SP, Tandan SK, Kumar D, Naik AK, Raviprakash V. (2007). Ameliorative effect of combined administration of inducible nitric oxide synthase inhibitor with

- cyclooxygenase-2 inhibitors in neuropathic pain in rats. *Eur J Pain*. 11:528-534.
8. Eisenberg E, McNicol ED, Carr DB. (2005). Efficacy and safety of opioid agonists in the treatment of neuropathic pain of nonmalignant origin. Systematic review and meta-analysis of randomized controlled trials. *JAMA*;293:3043-3052.
9. Eroglu L. (2002). Periferik Analjezikler. Ađrı, Ed. Erdine S. Abdi ibrahim Abdi. Algoloji Derneđi. İstanbul: 487-495.
10. Herrmann WA, Geertsens MS. (2009). Efficacy and safety of lornoxicam compared with placebo and diclofenac in acute sciatica/lumbo-sciatica: an analysis from a randomised, double-blind, multicentre, parallel-group study. *Int J Clin Pract*. 63:1613-1621.
11. Hu Y, Yang J, Hu Y, Wang Y, Li W. (2010). Amitriptyline rather than lornoxicam ameliorates neuropathic pain-induced deficits in abilities of spatial learning and memory. *Eur J Anaesthesiol*. 27:162-168.
12. Katz JA. (2000). Nonsteroidal anti-inflammatory analgesics. In: Raj PP. ed. Practical Management of Pain. Missouri: St Louis; p. 477-488.
13. Kimura S, Kontani H. (2009). Demonstration of antiallodynic effects of the cyclooxygenase-2 inhibitor meloxicam on established diabetic neuropathic pain in mice. *J Pharmacol Sci*. 110:213-217.
14. (2007). Martindale: The Complete Drug Reference, 35th Edition. Edited by Sean C Sweetman BPharm FRPharmS. Published by Pharmaceutical Press, London, UK.
15. McCormack K. (1994). Non-steroidal anti-inflammatory drugs and spinal nociceptive processes. *Pain*. 59:9-43.
16. Miranda HF, Puig MM, Dursteler C, Prieto JC, Pinardi G. (2007). Dexketoprofen induced antinociception in animal models of acute pain: synergy with morphine and paracetamol. *Neuropharmacology*. 52:291-6.
17. Miranda HF, Romero MA, Puig MM. (2012). Antinociceptive and anti-exudative synergism Miranda HF, Romero MA, Puig MM. Antinociceptive and anti-exudative synergism between dexketoprofen and tramadol in a model of inflammatory pain in mice. *Fundam Clin Pharmacol*. 26:373-382.
18. M ller KA., Johansson B. (1998). Odd-Geir Berge Assessing mechanical allodynia in the rat paw with a new electronic algometer, *Journal of Neuroscience Methods*. 84:41-47.
19. Namaka M, Gramlich CR, Ruhlen D. (2004). A treatment algorithm for neuropathic pain. *Clin Ther*. 26:951-956.
20. Namaka M, Leong C, Grossberndt A, Klowak M, Turcotte D. et al. (2009). A treatment algorithm for neuropathic pain: an update. *Consult Pharm*.24:885-902.
21. Raffa R, Pergolizzi JV, Tallarida RJ. (2010). Analgesic combinations. *J Pain*.11:701-704.
22. Steiner AA, Li S, Llanos- Q J, Blatteis CM. (2001). Differential inhibition by nimesulide of the early and late phases of intravenous and intracerebroventricular-LPS-induced fever in guinea pigs. *Neuroimmunomodulation*. 9:263-275.
23. Takahashi M, Kawaguchi M, Shimada K, Nakashima T, Furuya H. (2005). Systemic meloxicam reduces tactile allodynia development after L5 single spinal nerve injury in rats. *Reg Anesth Pain Med*. 30:351-5.
24. Treede RD, Jensen TS, Campbell JN, Cruccu G, Dostrovsky JO. et al. (2008). Neuropathic pain: redefinition and a grading system for clinical and research purposes. *Neurology*. 70:1630-1635.
25. Tuncer A, İp i Y, Aslantaş A, Ulug l A. (2003). N ropatik ađrı: semptomları, deneysel modelleri, patogenezi, tedavisi. İla  ve Tedavi Derg; 16:9-16.
26. Ulug l A. (2012). Ratlarda N ropati Modelleri. *Journal of Clinical and Analytical Medicine Kitap Serisi*;118-122:



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