

Bioactive Profiling of Essential oil of Terminalia Arjuna Stem Bark Collected from Orathur Village, Tamilnadu, India

Olujimi John Alagbe ^{1*} and Anorue, D.N ²

¹Department of Animal Nutrition and Biochemistry, Sumitra Research Institute, Gujarat, India.

²Department of Animal Science, University of Abuja, Nigeria.

Corresponding author: Olujimi John Alagbe, Department of Animal Nutrition and Biochemistry, Sumitra Research Institute, Gujarat, India.

Received date: December 13, 2024; **Accepted date:** January 10, 2025; **Published date:** January 29, 2025

Citation: Olujimi J. Alagbe and Anorue, D.N, (2025), Bioactive Profiling of Essential oil of Terminalia Arjuna Stem Bark Collected from Orathur Village, Tamilnadu, India, *J. Nutrition and Food Processing*, 8(1); DOI:[10.31579/2637-8914/287](https://doi.org/10.31579/2637-8914/287)

Copyright: © 2025, Olujimi John Alagbe. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract:

Bioactive profiling of *Terminalia arjuna* stem bark essential oil by GC/MS intends to showcase the medicinal properties and characterization of bioactive compounds. Bioactive profiling of essential oils from *Terminalia arjuna* stem bark revealed the presence of 31 bioactive compounds with their retention time. Cyclohexylhexanoate (10.78 %), D-limonene (9.57 %), ethyltrans-4-decenoate (9.52 %), α-himachalene (7.21 %), β-sesquiphellandrene (6.09 %), β-caryophyllene (5.66 %), Trans-2-Tetradecen-1-ol (4.09 %), β-Guaiene (4.02 %), 2-methyldecahydronaphthalene (3.72 %), cis-7-hexadecane (3.11 %), α-cadinol (3.04 %), 1-octanal (2.57 %) and ethylbenzene (2.02 %) were the major compounds above 2 % while compounds less than 2.0 % includes, 3-Hexenylhexanoate (0.97 %), 2,6,11-Trimethyldodecane (1.36 %), 2,3,6,7-Tetramethyloctane (0.25 %), β-Selinol (1.77 %), (-)-δ-Cadinol (0.01 %), Cubenol (0.03 %), α-Bisabolol (0.04 %), α-Himachalene (1.88 %), 1,3,5,8-Undecatetraene (1.02 %), Ethyltrans-4-Decenoate (0.05 %), α-Terpinolene (0.94 %), Trans-2-Nonenal (0.06 %), Geranyl Acetone (1.67 %), Cis-6-Pentadecen-1-ol (0.51 %) and Hexahydrofarnesol (0.87 %). It was concluded that essential oil from *Terminalia arjuna* stem bark is rich in several phytocomponents with medicinal properties and can be used to reduce the increasing cases of antimicrobial resistance.

Key words: *Terminalia arjuna*; phytocomponents; safety; medicine; antimicrobial; resistance

Introduction

Terminalia arjuna is an evergreen shrub from the Combretaceae family (Kapoor et al. 2014). The tree is distributed in India, Sri Lanka, China, Pakistan, Bangladesh, and Malaysia (Pashazanousi et al., 2012). The tree may grow up to 30 meters tall and is very therapeutic due to the presence of tannins, alkaloids, flavonoids, saponins, glycosides, and phenolic compounds, among other things (Saha et al., 2012; Bharani et al., 2004). These phyto-components have a variety of biological functions, including anti-inflammatory (Alagbe et al., 2021), antifungal, antiviral, antimicrobial, immune stimulator, cytotoxic, gastro-protective, anti-ulcer, anti-diabetic, hypolipidemic, antioxidant, osteogenic, anti-helminthic, and cardio-protective properties (Bharani et al., 2004; Paul et al., 2016). The plant parts (leaves, stem bark, and root extracts) have reportedly been used for the treatment of severe diarrhoea and dysentery, urethral discharge, gastro-intestinal infection, chest, pain, waist pain, irregular menstruation, internal pile, malarial, quick ejaculation, headache, hypertension, dysentery, premature aging, memory improvement, blood cleansing, chronic venous insufficiency, mental function, minor burns, scars, skin ulcers, varicose veins, wound healing.

The stem bark of the plant has sweet, cooling, styptic, tonic, anti-dysenteric, and febrifuge qualities (Desai et al., 2015). *Terminalia arjuna* leaf and root extracts have been shown to prevent the growth of pathogenic organisms such as *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Escherichia coli*, *Bacillus subtilis*, and *Candida albicans*. According to Singh et al. (2022; Alagbe, 2023), the concentration of phytocomponents in medicinal plants can be modified by several factors, including plant age, species, geographical location, and extraction process. These phyto-components have been shown to be safe, environmentally beneficial, and require no withdrawal period (Alagbe et al., 2020). However, errors in botanical identification, interference of medicinal plants with conventional pharmaceutical therapy, and a lack of studies on the adverse effects of medicinal plants can lead to toxicity in phytomedicine in humans and animals (Olujimi et al., 2024).

However, errors in botanical identification, Interference of medicinal plants and conventional pharmacological therapy and dearth of reports on

the side effects of medicinal plants can cause toxicity in phytomedicine in human and animals (Olujimi et al., 2024).

Therefore, this study was carried out to determine the bioactive profiling of essential oil of *Terminalia arjuna* stem bark collected from Orathur village, Tamilnadu, India

Materials and methods

Description of experimental area

The experiment was carried out at the department of Biochemistry, Sumitra Research Institute, Gujarat located between 28° 20' N and 75° 30' East India in the months of August to October, 2022.

Collection and extraction of essential oil from *Terminalia arjuna* stem bark

Fresh mature stem bark from *Terminalia arjuna* was collected from various areas in Orathur village, Tamil Nadu, India, and delivered to the taxonomy section of the same institute for proper authentication before being granted an identity number (HF/008C/2023). The essential oil was extracted from *Terminalia arjuna* stem bark using the steam distillation process with the aid of Clevenger apparatus. The extracted oil was forwarded to the laboratory for further investigation.

Bioactive profiling of essential oil from *Terminalia arjuna* stem bark

Bioactive profiling of essential oil from *Terminalia arjuna* stem bark was carried out using Lauret gas chromatography - mass spectrometry (Model FG/008, Netherlands). Identification of each bioactive compound was carried out by comparing their mass spectra with those of reference compounds from the Library of National Institute of Standard and Technology (NIST, 2011) database.

S/N	Compounds	Reaction time (min)	% Area
1	3-Hexenylhexanoate	5.62	0.97
2	2,6,11-Trimethyldodecane	6.27	1.36
3	β-Caryophyllene	6.33	5.66
4	Cyclohexylhexanoate	7.07	10.78
5	γ-Cadinene	7.55	2.67
6	β-Sesquiphellandrene	7.92	6.09
7	β-Linalool	8.09	2.51
8	D-Limonene	8.47	9.57
9	2,3,6,7-Tetramethyloctane	8.84	0.25
10	β-Selinol	8.93	1.77
11	α-Cadinol	9.62	3.04
12	(-)δ-Cadinol	9.95	0.01
13	α-Bisabolol	10.50	0.04
14	Cubenol	11.10	0.03
15	α-Himachalene	11.55	7.21
16	β-Guaiene	12.35	4.02
17	α-Himachalene	12.67	1.88
18	1,3,5,8-Undecatetraene	12.85	1.02
19	Ethyltrans-4-Decenoate	13.06	0.05
20	α-Terpinolene	14.54	0.94
21	1-Octanal	15.12	2.57
22	1,8-Cineole	15.76	3.5
23	Ethyltrans-4-Decenoate	16.27	9.52
24	2-methyldecahydronaphthalene	17.16	3.72
25	Ethylbenzene	18.09	2.02
26	Trans-2-Nonenal	19.22	0.06
27	Geranyl Acetone	19.85	1.67
28	Cis-6-Pentadecen-1-ol	20.06	0.51
29	Trans-2-Tetradecen-1-ol	21.38	4.09
30	Cis-7-Hexadecane	22.40	3.11
31	Hexahydrofarnesol	22.75	0.87
Total		91.51	
Number of compounds			
Monoterpenes		27.51	
Diterpenes		7.96	
Triterpenes		1.03	
Sesquiterpenes		-	
Non-terpenes		55.01	

Table 1: Bioactive profiling of *Terminalia arjuna* stem bark essential oil by GC/MS

Results and Discussion

Bioactive profiling of essential oils from *Terminalia arjuna* stem bark identified 31 bioactive components and their retention times.

Cyclohexylhexanoate (10.78%), D-limonene (9.57%), ethyltrans-4-decenoate (9.52%), α-himachalene (7.21%), β-sesquiphellandrene (6.09%), β-caryophyllene (5.66%), Trans-2-Tetradecen-1-ol (4.09%), β-Guaiene (4.02%), 2-methyldecahydronaphthalene (3.72%), cis-7-

hexadecane (3.11%), α -cadinol (3.04%). It is worth noting that all of these bioactive molecules, often known as phytochemicals, have medical or therapeutic characteristics. This result is consistent with prior research by Kokkiripati et al. (2013), Hafiz et al. (2014), and Chaudhari and Mengi (2006). Cyclohexylhexanoate, β -caryophyllene, β -Linalool, β -sesquiphellandrene, β -selinenol, and α -cadinol have been shown to have antimicrobial, antifungal, antidiarrhea, antibacterial, anticancer, antioxidant, and anti-helminthic properties (Subavathy and Thilaga, 2015; Mangrove et al., 2014). Doughari (2012) and Olajuyige et al. (2011) found that 2, 6, 11-trimethyldodecane, 2, 3, 6, 7-tetramethyloctane, α -himachalene, α -terpinolene, and cis-6-pentadecen-1-ol have antibacterial and gastro-protective properties. α -bisabolol, 2-methyldecahydronaphthalene, and ethyltrans-4-deenoate have been shown to have antibacterial and cardio-protective effects (Devendran and Baasubramanian, 2011; Lima et al., 2010). Trans-2-nonenal, geranyl acetone and ethylbenzene have antifungal and anti-diarrhoea properties (Mamza et al., 2012; Awa et al., 2012). Screening for bioactive chemicals in herbal plants can lead to the development of new medical medicines with effective disease prevention and treatment properties (Soma et al., 2010; Alagbe et al., 2024). The concentrations of phytochemicals in herbal plants can be altered by several factors, including plant age, geographical location, species, and processing methods (Alagbe et al., 2023a; Alagbe et al., 2023b).

Conclusion

Naturally, medicinal plants contain phytochemicals with therapeutic effects. These chemicals have a wide range of biological functions, including antibacterial, antifungal, antihelminthic, hepatoprotective, immune-stimulatory, cytotoxic, antioxidant, and antiviral properties.

References

1. Alagbe, J.O (2024).Proximate, mineral and phytochemical analysis of some medicinal plants collected from Orathur village, Thirupur Taluk Kancheepuram district Tamilnadu, India. *World Journal of Agriculture and Forestry Sciences*, 2(2): 30-35.
2. Mamza, U.T., Sodipo, O.A. & Khan, I.Z. (2012). Gas Chromatography - Mass spectrometry (GC - MS) analysis of bioactive components of *Phyllanthus amarus* Leaves. International Research Journal of Plant Science, 3 (10), 208 - 215
3. Awa, E.P., Ibrahim, S. & Ameh, D.A. (2012). GC/MS Analysis and Antimicrobial Activity of Dimethyl Ether fraction of Methanolic Extract from stem Bark of *Annona senegalensis* Pers. International Journal of Phamaceutical Sciences and Research. 3 (11), 4213-4218
4. Devendran, G. & Balasubramanian, U. (2011). Qualitative phytochemical screening and GC-MS analysis of *Ocimum Sanctum L.* leaves. Asian Journal of Plant Science and Research, 1 (4), 44- 48.
5. Lima, A.L., Parial, R., Das, M. & Das, A.K. (2010). Phytochemical and Pharmacological studies of ethanolic extact from the leaf of mangrove plant *Phoenix paludosa* Roxb. Malaysian Journal of Pharmaceutical Sciences, 8 (2), 59-69.
6. Doughari, J.H. (2012). Phytochemicals: Extraction Methods, Basic Structures and Mode of Action as Potential Chemmoterapeutic Agents- A Global Perspective of Their Role in Nutrition and Health. Dr. Venketeshwer Rao. Ed. ISBN: 978- 953-51-0296-0. Intech.
7. Olajuyige, O.O., Babalola, A.E. & Afolalayan, A.J. (2011). Antibacterial and phytochemical screening of crude ethanolic extracts of *Waltheria Linn*. African Journal of Microbiology Research, 5 (22), 3760-3764.
8. Subavathy, P. & Thilaga, R.D. (2015). GC-MS Analysis of Bioactive Compounds from Whole Body of Methanolic Extract of *Cypraea arabica*. World Journal of Pharmaceutical Research, 5 (3), 800-806.
9. Mangrove-Abayomi, O.E., Kenneth, E and Mkaparu, K.I. (2014). Chemometric profiling of methanolic leaf extract of *Cinddoscolus aconitifolius* (Euphorbiaceae) using UV-VIS, FTIR and GC-MS techniques. Peak journal of Medicinal Plant Research, 2 (1), 6-12.
10. Alagbe, J.O., Bamigboye, S., Nwosu, G.C., Agbonika, D.A and Kadiri Mercy Cincinsoko. (2023a). Characterization of bioactive compounds in *Luffa aegyptiaca* leaf ethanolic extracts using gas chromatography and mass spectrometry (GC-MS). *Drug Discovery*, 2023; 17:e10dd1011.
11. Alagbe, J.O., Kadiri, M.C., Oluwafemi, R.A., Agubosi, O.C.P and Anorue, D.N. (2023b). Analysis of bioactive compounds in ethanolic extracts of *Xylopia aethiopica* leaves using gas chromatography and mass spectrometry technique. *American Journal of Science on Integration and Human Development*, 1(1): 1-10.
12. Alagbe, J.O. (2023). Bioactive compounds in ethanolic extract of *Strychnos innocua* root using gas chromatography and mass spectrometry (GC-MS). *Drug Discovery*, 2023; 17:e4dd1005.
13. Singh Sharma., Alagbe Olujimi John., Liu Xing., Sharma Ram and Kumar Amita (2022). Comparative analysis of ethanolic *Juniperus thurifera* leaf, stem bark and root extract using gas chromatography and mass spectroometry. *International Journal of Agriculture and Animal Production*, 2(6): 18-27.
14. Alagbe, J.O., Adedeji, M.O., Habiba, Z., Nwosu, Gloria and Wyedia Dabara Comfort (2021). Physico-chemical properties of *Indigofera zollingeriana* seed oil. *Asian Journal of Advances in Medical Science* 3(4): 306-308.
15. Alagbe, J.O., Shittu, M.D and Ushie, F.T. (2021). GC-MS analysis of methanolic stem bark extract of *Zollingeriana indigofera*. *Asian Journal of Advances in Research* 11(4): 144-146.
16. Alagbe, J.O (2020). Chemical evaluation of proximate, vitamin and amino acid profile of leaf, stem bark and roots of *Indigofera tinctoria*. *International Journal on Integrated Education*. 3(10): 150-157.
17. Shittu, M.D and Alagbe, J.O. (2020). Phyto-nutritional profiles of broom weed (*Sida acuta*) leaf extract. *International Journal of Integrated Education*. 3(11): 119-124
18. Kapoor D, Vijayvergiya R, Dhawan V. (2014). Terminalia arjuna in coronary artery disease: Ethnopharmacology, pre-clinical, clinical and safety evaluation. *Journal of Ethnopharmacology*, 155:1029-1045.
19. Pashazanousi MB, Raeesi M, Shirali S. (2012). Chemical composition of the essential oil, antibacterial and antioxidant

- activities, total phenolic and flavonoid evaluation of various extracts from leaves and fruit peels of Citrus limon. Asian Journal of Chemistry, 24:4331-4334.
20. Hafiz FB, Towfique NM, Sen MK, Sima SN, Azhar BS, Rahman MM. (2014). A comprehensive ethnopharmacological and phytochemical update review on medicinal plant of Terminalia arjuna Roxb. of bangladesh. Sch Acad J Pharmacology, 3:19-25
 21. Chaudhari M, Mengi S. (2006). Evaluation of phytoconstituents of Terminalia arjuna for wound healing activity in rats. Phytother Research, 20:799-805.
 22. Kokkiripati PK, Kamsala RV, Bashyam L, Manthapuram N, Bitla P, Peddada V. (2013). Stem-bark of Terminalia arjuna attenuates human monocytic (THP-1) and aortic endothelial cell activation. J Ethnopharmacology, 146:456-464
 23. Olujimi, J.O., Anuore, D.N and Aliyu, K.I. (2024). Chrysophyllum albidum stem bark powder: effects on performance and carcass characteristics of Japanese quails. World Journal of Clinical Studies, 2(2): 41-48.
 24. Saha A, Pawar VM, Jayaraman S. (2012). Characterization of polyphenols in Terminalia arjuna bark extract. Indian J Pharm Science, 74:339-347
 25. Bharani A, Ahirwal K, Jain N. (2004). Terminalia arjuna reverses impaired endothelial function in chronic smokers. Indian Heart Journal, 56:123-128.
 26. Paul S, Ghosh D, Ghosh AK, Bhowmick D, Bandyopadhyay D, Chattopadhyay A. (2016). Aqueous bark extract of Terminalia arjuna protects against phenylhydrazine induced oxidative damage in goat red blood cell membrane bound and metabolic enzymes. Int J Pharm Pharm Science, 8:62-70.
 27. Khalil S. (2005). Effect of Statin Versus Terminalia arjuna on Acute Myocardial Infarction (DNB thesis). New Delhi, India: National Board of Examination; 2005.
 28. Desai SD, Patil BS, Kanthe PS, Potekar RM. (2015). Effect of ethanolic extract of Terminalia arjuna on liver functions and histopathology of liver in albino rats fed with hyperlipidemic diet. Int J Pharm Pharm Science, 7:302-306.
 29. Bharani A, Ganguli A, Bhargava KD. (1995). Salutary effect of Terminalia arjuna in patients with severe refractory heart failure. Int J Cardiology, 49:191-199.
 30. Yaidikar L and Thakur S. (2015). Arjunolic acid, a pentacyclic triterpenoidal saponin of Terminalia arjuna bark protects neurons from oxidative stress associated damage in focal cerebral ischemia and reperfusion. Pharmacol Repository, 67:890-895.



This work is licensed under Creative Commons Attribution 4.0 License

To Submit Your Article Click Here:

[Submit Manuscript](#)

DOI:[10.31579/2637-8914/287](https://doi.org/10.31579/2637-8914/287)

Ready to submit your research? Choose Auctores and benefit from:

- fast, convenient online submission
- rigorous peer review by experienced research in your field
- rapid publication on acceptance
- authors retain copyrights
- unique DOI for all articles
- immediate, unrestricted online access

At Auctores, research is always in progress.

Learn more <https://auctoresonline.org/journals/nutrition-and-food-processing>