Preimplantation Genetic Testing (PGT) for Non-Lethal Correctable Conditions Through Population Carrier Screening

Svetlana Rechitsky¹, Tatiana Pakhalchuk¹, Maria Prokopovich¹, Agnes Machaj¹, Anver Kuliev^{1*}

1 Reproductive Genetics Innovations, Northbrook, Illinois.

*Corresponding Author: Anver Kuliev, Reproductive Genetics Innovations, Northbrook, Illinois.

Received date: August 11, 2023; Accepted date: August 29, 2023; Published date: September 07, 2023.

Citation: Svetlana Rechitsky, Tatiana Pakhalchuk, Maria Prokopovich, Agnes Machaj, Anver Kuliev (2023), Preimplantation Genetic Testing (PGT) for Non-Lethal Correctable Conditions Through Population Carrier Screening, *J. Obstetrics Gynecology and Reproductive Sciences*, 7(6) **DOI:**10.31579/2578-8965/182

Copyright: © 2023, Anver Kuliev. This is an open-access article distributed under the terms of The Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract:

An increasing number of patients without family history of genetic disease are referred for preimplantation genetic testing of monogenic disorders (PGT-M) after ascertainment through population (expanded) carrier screening (ECS), increasing the proportion of non-lethal correctable genetic conditions in PGT-M indication profile. The aim of this paper is to analyze the current dynamics of these changes in our PGT-M series, which is the world's largest PGT-M experience. This was analyzed using our PGT-M for two prevalent examples of non-lethal correctable genetic conditions, phenylketonuria (PKU) and hereditary hearing loss (HHL) during the period of 2005-2022. During this period, 91 PGT-M cycles were performed for PKU, involving testing for 108 mutant alleles in PAH gene, and 262 PGT-M cycles for HHL, involving testing for up to 300 mutations in 13 different genes causing hearing loss. PGT-M for these non-lethal correctable conditions, with 87% of prospective PGT-M performed for PKU, and 82% for HHL, performed for couples who had no affected relatives with PKU or HHL. These at-risk couples were ascertained through ECS in the last decade, making it possible to provide the option of prospective PGT-M for these non-lethal correctable conditions.

Keywords: preimplantation genetic testing for monogenic disorders (pgt-m); phenylketonuria (pku); hereditary hearing loss (hhl); prospective application of pgt-m, expanded carrier screening (ecs), non-lethal correctable conditions

Introduction

Although phenylketonuria (PKU) and hereditary hearing loss (HHL) (https://hereditaryhearingloss.org/) are non-lethal conditions, they still require lifelong care, representing a real financial burden. Prenatal diagnosis of these conditions is also possible but leads to a difficult decision of a possible termination of affected pregnancy, which is the problem for any non-lethal condition. Preimplantation genetic testing (PGT), based on preselection and transfer of unaffected embryos avoids the risk for pregnancy termination, and most importantly, allows having children free of these conditions.

Since the introduction of PGT for monogenic disorders (PGT-M) [1,2], it has been applied for more than 600 different conditions, resulting in birth of thousands of children free of genetic disease [3]. Initially applied for avoiding a recurrence risk of having severe genetic conditions, PGT-M has

then been applied also for diseases with genetic predisposition, such as breast cancer and coronary disease [4-7]. It has been usually applied retrospectively for those at-risk couples who have already had an affected offspring with monogenic disorder, until introduction of different screening programs, including expanded (population) carrier screening (ECS), which made it possible to apply PGT-M for patients without an affected family member, with a chance of avoiding any possibility of having an affected child in the family. Accordingly, an increasing number of patients without family history of genetic disease are referred for PGT-M after ascertainment through ECS, also significantly increasing the proportion of non-lethal correctable genetic conditions in the indication profile. The aim of this paper is to analyze the current dynamics of these changes, based on our PGT-M series, which is the world's largest PGT-M experience. As will be presented, using PGT-M for PKU and HHL, the examples of non-lethal correctable conditions, there is an ongoing shift of PGT-M from retrospective to prospective application, as a practical tool for primary prevention of genetic disorders.

Materials and Methods

Dynamics of referrals for PGT-M of PKU and HL were analyzed during the period of 2005-2022. These included 91 PGT-M cycles for PKU, performed for 53 at risk carriers of 108 mutant alleles in PAH gene, including 98

J. Obstetrics Gynecology and Reproductive Sciences

Copy rights @ Anver Kuliev, et al

missence and 12 spicing mutations, and 1 deletion. For ten of these carries, a combined PGT-M was performed for additional conditions, including cystic fibrosis, fragile-X mental retardation, BRCA-1, myotonic dystrophy, adrenal hypoplasia, Wilson's disease, propionic acidemia and translocation.

PGT-M for HHL was done in 262 PGT-M cycles for 178 patients at risk for producing progeny with syndromic and non-syndromic HHL, caused by up to 300 mutations in 13 different genes (Table 1).

-			
Gene	Omim	Patients	Cycles
PLCB4	614669	1	3
GJB6	612643	1	2
MYO15 A	600316	1	1
TMPRS S3	601072	2	3
GJB2	220290	140	201
PRPS1	304500	1	1
SLC26A 4	274600	7	10
MYO7A	276900	4	4
PCDH1 5	602083	3	5
USH2A	276901	13	21
ADGRV 1	605472	1	2
MITF	193510	3	7
WFS1	222300	1	2
(13 genes)		178	262
	Gene PLCB4 GJB6 MYO15 A TMPRS S3 GJB2 PRPS1 SLC26A 4 MYO7A PCDH1 5 USH2A ADGRV 1 MITF WFS1 (13 genes)	Gene Omim PLCB4 614669 GJB6 612643 MYO15 600316 TMPRS 601072 S3 601072 GJB2 220290 PRPS1 304500 SLC26A 274600 MYO7A 276900 PCDH1 602083 USH2A 276901 ADGRV 605472 MITF 193510 WFS1 222300 (13 (13 genes)	Gene Omim Patients PLCB4 614669 1 GJB6 612643 1 MYO15 600316 1 TMPRS 601072 2 GJB2 220290 140 PRPS1 304500 1 SLC26A 274600 7 MYO7A 276900 4 PCDH1 602083 3 USH2A 276901 13 ADGRV 605472 1 MITF 193510 3 WFS1 222300 1 (13 genes) 178

 Table 1: List of Syndromic and Non-syndromic HHL for which PGT-Mwas performed during 2005-2022

Initially, most of these patients tested for PKU and HHL were referred for avoiding a recurrence of these conditions after birth of an affected child in the family, but the number of such patients has been decreasing in the last decade, with a corresponding increase of couples with no family history of these conditions, who referred for PGT-M to avoid even the first occurrence of the disease.

PGT-M methods applied during this period have changed from traditional mutation testing with linkage analysis using informative short tandem repeats (STRs), to more sophisticated approaches allowing a combined testing for single gene and chromosomal disorders [8]. In brief, PGT-M was performed using a standard IVF protocol, coupled with embryo biopsy, described in detail elsewhere [9]. Details of PGT guidelines were reported previously [10, 11]. The present standards of the procedure involve a whole genome amplification (WGA) of embryo samples obtained by blastocyst

biopsy, followed by multiplex nested PCR analysis of the mutations in question, together with closely linked genetic markers in a multiplex heminested system [8,9]. For each family, heterozygous alleles and haplotypes not shared by parents were selected. This allowed detecting and avoiding misdiagnosis due to preferential amplification and allele dropout (ADO), and a possible aneuploidy or uniparental disomy of chromosomes in which the tested mutations are located, which may affect diagnostic accuracy of PGT-M. In PGT cycles, involving an advanced reproductive age of maternal partner, aneuploidy testing was also performed by next generation technologies (NGS) (Illumina Inc) for 24-chromosome aneuploidy analysis [8].

Results and Discussion

Results of 91 PGT-M cycles for PKU are presented in Table 2,

Disease	Gene	Patients	Cycles	Transfers	Embryos transferred	Pregnancy	Babies
PHENYLKETON URIA (PKU)	РАН	53	91	111	129	53 48%	54

Table 2: PGT-M for Phenylketonuria (PKU)

J. Obstetrics Gynecology and Reproductive Sciences

showing the detection of 129 of PKU free embryos for transfer in 111 cycles, yielding 53 (48%) clinical pregnancies and birth of 54 unaffected babies. PGT-M referrals during the period of 2005-2015 were mainly (75%) after the birth of affected child, while after 2016 only a few (13%) PGT-M referral were due to an affected progeny in the family, while the majority (87%) were

performed prospectively for couples who had no affected relative but were ascertained through ECS.

Similar results were observed in the dynamics of 262 PGT-M cycles performed for HHL, which resulted in transfer of 282 HHL free embryos in 258 cycles, yielding 164 (63.5%) clinical pregnancies and birth of 134 children free of HHL (Table 3).

Number of Genes	Patients	Cycles	Transfers	Embryos Transferred	Pregnancy	Babies
13	178	262	258	282	164 (63.5%)	134

Table 3: PGT-M for Hereditary Hearing Loss (HHL)

PGT-M referrals during the period of 17 years showed the shift from retrospective to prospective application of PGT-M, with 82% -provided for couples with no affected relative, who were ascertained through ECS in the last decade (Table 4).

Years	Total Patients presented for PGT _f M	With Affected relative	Without Affected Relative	
2005-2015	21	15	6 (40%)	
2016-2022	196	36	160 (82%)	

 Table 4: Increase of PGT-M cases for Hereditary Hearing Loss (HHL) with no affected relative in the family, ascertained through expanded carrier screening (ECS)

The data show an increase of referral for PGT-M of PKU and HHL, which originally have not even been an indication for PGT-M. Such an increase may be related to the introduction of ECS, which allows couples at risk avoiding even the first occurrence of disease. Retrospective application of PGT-M was mainly prior-to 2016, performed to prevent a recurrence of PKU or HHL after couples have had an affected offspring or had a relative with these conditions. However, increasing number of patients after 2015 presented with no relative affected with PKU or HHL. Overall, 87% PGT-M cycles for PKU and 82% for HHL were performed prospectively for couples with no affected relative.

With a low treatment efficiency for therapeutic interventions and a high (25% - 50%) risks of recurrence in each pregnancy, PGT-M may be the main tool for the purpose of primary prevention of genetic disease. However, atrisk couples usually do not know about their at-risk status until they have an affected child, so reduction of affected birth prevalence cannot be done in the absence of screening programs, which have been offered for many years exclusively for special ethnic groups or selected genetic conditions. But in contrast to such screening programs applied to special groups of high incidences, ECS is offered in pan-ethnic fashion, extending surveillance to couples of all ethnicities and involving increasing number of genetic conditions [12,13], that appeared having a growing impact on PGT-M application. Targeted gene panels further increase the number of genetic disorders amenable to ECS, leading to PGT-M prospectively, representing a real form of primary prevention, as it allows avoiding affected pregnancies, and establishing unaffected pregnancies from the onset. Several hundred additional genes are becoming considered plausible candidates for inclusion into ECS, with expanded panels now consisting of hundreds of genes, resulting in the increase of detection rate [14]. Thus, presented dynamics of PGT-M during the period of 17 years demonstrated the shift from retrospective to prospective application of PGT -M for PKU and HHL. Overall, 87% and 82% PGT cycles for PKU and HHL, respectively, were performed prospectively for couples with no affected relative. An increase in referrals for PGT-M through ECS was observed also for other genes, with the number of prospective PGT-M cases for the last decade more than doubling through referral from ECS [15]. This may lead to the application PGT-M beyond family level, allowing to offer PGT-M prospectively before the birth of an affected child, as a tool for primary prevention of genetic disorders.

References

- Handyside AH, Kontogianni EH, Hardy K, et al. (1990). Pregnancies from biopsied human preimplantation embryos sexed by Y-specific DNA amplification. Nature 344(6268):768-770.
- Verlinsky Y, Ginsberg N, Lifchez A, Valle J, MoiseJ, Strom CM (1990). Analysis of the first polar body: Preconception genetic diagnosis. Hum Reprod 5: 826-829.
- Kuliev A, Rechitsky S, Simpson JL (2020). Practical preimplantation genetic testing. (3rd edn) Springer Nature, Switzerland.
- 4. Rechitsky S, Verlinsky O, Chistokhina A, Sharapova T, Masciangelo C, et al. (2002). Preimplantation genetic diagnosis for cancer predisposition. Reprod Biomed Online. 4:148-155.
- Kuliev A, Pakhalchuk T, Prokhorovich M, Rechitsky S (2021). Preimplantation Genetic Testing for Inherited Predisposition to Cardiac Disease. Ann Heart 5(1):96-99.

J. Obstetrics Gynecology and Reproductive Sciences

- 6. Vriesen N, Carmany EP, Natoli JL (2022). Clinical outcomes of preimplantation genetic testing for hereditary cancer syndromes: a systematic review. Prenat Diagn 42(2):201-211
- Rechitsky S., Pakhalchuk T., Prokhorovich M., Magal A, Kuliev A. (2023). Expanding Application of Preimplantation Genetic Testing (PGT) for *Breast Cancer. J. Obstetrics Gynecology and Reproductive Sciences*; 7(4).
- Rechitsky S, Pakhalchuk T, San Ramos G, Goodman A, Zlatopolsky Z, et al. (2015). First systematic experience of preimplantation genetic diagnosis for single-gene disorders, and/or preimplantation human leukocyte antigen typing, combined with 24-chromosome aneuploidy testing. Fertil Steril 103:503-512.
- 9. Kuliev A, Rechitsky S, Verlinsky O (2014). Atlas of preimplantation genetic diagnosis. An Illustrated Textbook. (3rd ed.), Taylor & Francis, London.
- Preimplantation Genetic Diagnosis International Society (PGDIS) (2008). Guidelines for good practice in PGD: Program requirements and laboratory quality assurance. *Reprod BioMed Online* 16: 134-147.

Copy rights @ Anver Kuliev, et al

- 11. ESHRE PGD Consortium Steering Committee (2020). ESHRE PGD consortium good practice recommendations for the organization of PGT. Human Reproduction Open, 29;2020(3): hoaa021.
- 12. Edwards JG, Feldman G, Goldberg J, et al (2015). Expanded carrier screening in reproductive medicine-points to consider. *Obstet Gynecol*.125(3):653-662.
- Haque IS, Lazarin GA, Kang P, Evans EA, Goldberg JD, Wapner RJ (2016). Modeled fetal risk of genetic diseases identified by expanded carrier screening. JAMA, 3016(7):734-742.
- Hogan GJ, Vysotskaia VS, Beauchamp KA, Seisenberger S, Grauman PV, et al (2018). Validation of an Expanded Carrier Screen that optimizes sensitivity via full-exon sequencing and panel-wide copy number variant identification. Clin Chem 64:1063-1073.
- 15. Simpson JL, Rechitsky S, Kuliev A (2019). Before the beginning: The genetic risk of a couple aiming to conceive Fertil Steril 112:622-630.



This work is licensed under Creative Commons Attribution 4.0 License

To Submit Your Article, Click Here:

Submit Manuscript

DOI:10.31579/2578-8965/182

Ready to submit your research? Choose Auctores and benefit from:

- ➢ fast, convenient online submission
- > rigorous peer review by experienced research in your field
- rapid publication on acceptance
- > authors retain copyrights
- > unique DOI for all articles
- immediate, unrestricted online access

At Auctores, research is always in progress.

Learn more <u>https://www.auctoresonline.org/journals/obstetrics-gynecology-and-reproductive-sciences</u>