Central Nervous System Toxicity during the Induction of General Anesthesia After Intravenous Bolus of Lidocaine

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Abstract

Lidocaine is a frequently utilized intravenous adjuvant for the induction of general anesthesia. Despite its wide therapeutic margin of safety, central nervous system (CNS) and cardiovascular toxicity may ensue. Vigilance is essential in the recognition and immediate treatment of suspected systemic lidocaine toxicity. We present a case of CNS toxicity after an intravenous bolus of lidocaine during the induction of general anesthesia.

Objectives

- Summarize a description of the case including: the patient’s profile, the patient’s medications, the anesthesia management, the clinical presentation of lidocaine toxicity, and the management of lidocaine toxicity.
- Summarize the pharmacology, structure and mechanism of lidocaine.
- Understand the mechanism, recognition, and management of systemic lidocaine toxicity.
- Discuss the potential contributors to the development of lidocaine toxicity.
- Discuss novel practice recommendations.

Key Words: lidocaine; central nervous system (cns); cardiovascular toxicity

Introduction

Lidocaine is a frequently utilized intravenous adjuvant for the induction of general anesthesia. Despite its wide therapeutic margin of safety, central nervous system (CNS) and cardiovascular toxicity may ensue. Vigilance is essential in the recognition and immediate treatment of suspected systemic lidocaine toxicity. We present a case of CNS toxicity after an intravenous bolus of lidocaine during the induction of general anesthesia.

Case Presentation

A 22-year-old female presented for a temporomandibular joint (TMJ) arthroscopy and manipulation. The patient’s pertinent medical history included orofacial pain and daily migraines refractory to other treatment modalities including thoracenteses, physical therapy and use of a night guard. The patient had a past medical history significant for Chiari 1 malformation, intractable migraines with aura, tethered spinal cord, seasonal allergies, mild asthma, gastroesophageal reflux, MVA, and post-operative nausea and vomiting.

The patient’s home medication list included: acetaminophen 325 mg tablet, albuterol 90 mcg/actuation inhaler, almotriptan 12.5 MG tablet, aspirin-acetaminophen-coffeine 250-250-65 mg per tablet, botulinum toxin type A 100 unit SoSR, cetirizine 10 MG tablet, fluticasone propionate 50 mcg/actuation nasal spray, ibuprofen 200 MG tablet, levonorgestrel 14 mcg/24 hour, melatonin 10 mg tablet, oxycodone 5 MG tablet, and tramadol 50 mg tablet. Prior to the induction of anesthesia, the patient received 20 mg of intravenous famotidine, a transdermal 1.5 mg scopolamine patch, and an oral 975 mg dosage of acetyaminophen.

During induction of anesthesia, the patient developed anxiety, tinnitus, a foul taste in the mouth, and tremors immediately after receiving a dose of lidocaine at a 2 mg kg⁻¹. Patient was then given 2 mg morphine and 150 mg propofol followed by a prophylactic 2 mg intravenous bolus of midazolam. Subsequently, the patient received 80 mg of succinylcholine for intubation. A 6.0 mm Nasal Ra tube was successfully placed and confirmed via fiberoptic bronchoscopy. The patient was administered an FiO2 of 100%, inhaled volatile anesthetic (sevoflurane) and hyperventilated to an ETCO2 of 30 mmHg. The notable hemodynamic changes after intubation were tachycardia (HR=167 BPM) and hypertension (BP=150/85; MAP=106mmHg), which were corrected with a 10 mg bolus of IV esmolol. The patient’s ventilation was maintained with a target ETCO2 of 30-32 mmHg throughout the case.

The patient’s cardiovascular status remained stable throughout the duration of the procedure. The patient received no further administration of lidocaine by the surgical or anesthesia team. There were no post-operative complications.

Lidocaine: Pharmacology and Clinical Indications

The patient was administered 0.06 mg/kg IV Lidocaine followed by a 3 mg/kg IV bolus of lidocaine. The patient developed hypoxia (SaO2=90%) and hypotension (BP=60/40). The patient was treated with 100% O2, 2 mg morphine, and 1 mg intravenous atropine. The patient’s cardiovascular status remained stable.

The patient was then intubated and maintained with a target ETCO2 of 30-32 mmHg throughout the case. The patient received no further administration of lidocaine by the surgical or anesthesia team. There were no post-operative complications.
Lidocaine is a frequently utilized intravenous adjuvant for the induction of general anesthesia. Lidocaine’s molecular structure consists of a hydrophilic diethylglycine molecule bound via an amide bond to a hydrophobic aromatic benzene ring. The amphipathic nature of lidocaine permits permeability across both hydrophobic and hydrophilic membranes. Lidocaine metabolism is hepatic, via cytochrome P450 3A4. Lidocaine elimination half-life is 1.5-2 hour with renal excretion. The molecular structure of lidocaine (2-diethylaminoaceto-2',6'-xylidide) is displayed below:

Sodium channel antagonists, such as lidocaine, inhibit ectopic discharges of TTX-sensitive and TTX-resistant sodium channels without affecting nerve conduction. Sodium channel blockade results in a reduction of neuronal action potential initiation, propagation and facilitates membrane stabilization. The peak plasma concentrations (Cmax) obtained from a 1.5-2 mg/kg bolus of intravenous lidocaine inhibit a small number of sodium channels, thus secondary and tertiary mechanisms are likely involved. In progressively higher serum concentrations, lidocaine will inhibit voltage-gated potassium channels, voltage-gated calcium channels and nicotinic acetylcholine (nACh) receptors.

Intravenous lidocaine has been shown to target polymorphonuclear granulocytes by interfering with upstream G-protein pro-inflammatory signaling and the release of reactive oxygen species and cytokines. Intravenous lidocaine results in a reduction of cerebral metabolic rate, cerebral blood flow, and cerebral metabolic oxygen consumption, thus, reducing energy required for neuronal membrane stabilization. The CNS inhibitory effects of systemic lidocaine may be influenced by lidocaine mediated strychnine-sensitive glycine receptors of wide dynamic range neurons in the spinal cord. Lidocaine is also shown to block the K+-Cl− cotransporter (KCC) on γ-aminobutyric acid (GABA_B) receptors. This blockade and lidocaine mediated inhibition likely plays a role in the progressive nervous system symptoms and seizure activity observed in systemic lidocaine CNS toxicity.

N-methyl-D-aspartate (NMDA) receptors play an important role in nociception via fast excitatory neurotransmission in the CNS. Lidocaine plays a role in reducing hyperalgesia by its effects on NR1/2A NMDA receptor subunits. Lidocaine is shown to indirectly inhibit NFMDA via its effects on protein kinase C (PKC) in a concentration dependent manner. Lidocaine inhibits PKC mediated functions intracellularly, resulting in a reduction of PKC phosphorylation of NMDA subunits, consequently, reducing downstream signaling and NMDA receptor function.

A 1.5 mg kg⁻¹ bolus of intravenous lidocaine during anesthetic induction serves a number of purposes: the attenuation of pain response to propofol and/or etomidate administration, blunting the cough reflex elicited by a rapid fentanyl bolus and airway instrumentation, reduction of succinylcholine-induced elevations in intragastric pressure and ICP, mitigating the hemodynamic response (hypertension and tachycardia) from laryngoscopy, blunting increases in ICP secondary to airway instrumentation, and to mitigate airway reactivity. Systemic lidocaine has also been utilized for its analgesic, anti-inflammatory properties, and its synergistic reduction in the minimum alveolar concentration (MAC) of volatile anesthetics by 10-28%. Factors that may influence the unbound free fraction of systemic lidocaine include: a reduction in plasma protein binding, increased total interstitial volume, and reduced total plasma volume. Lidocaine’s high hepatic extraction ratio (0.63) makes it dependent on hepatic blood flow. The metabolism of lidocaine is dependent upon CYP450 3A4 enzyme function and hepatocellular perfusion. The hepatic metabolism of lidocaine occurs via CYP450 3A4 enzyme N-dealkylation and hydroxylation. The CYP450 3A4 metabolism results in the formation of monoethyglycinexilide via hepatic oxidative dealkylation and subsequently the formation of glycine xylidine via hydroxylation. The elimination half-life (t 1/2β) of lidocaine is 1.5-2 hours and excretion of metabolites occurs via the kidney with <5% excreted unmetabolized.

### Lidocaine: Selected Mechanisms of ACTION

<table>
<thead>
<tr>
<th>Target</th>
<th>Mechanism of Action</th>
</tr>
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<tbody>
<tr>
<td>Na+ channel antagonist</td>
<td>↓ ectopic discharges of TTX-sensitive and TTX-resistant Na+ channels</td>
</tr>
<tr>
<td>Potassium channel antagonist</td>
<td>Blocks voltage-gated K+ channels and nACh receptors</td>
</tr>
<tr>
<td>Calcium channel antagonist</td>
<td>Interference with upstream Gq-protein pro-inflammatory signaling</td>
</tr>
<tr>
<td>Nicotinic acetylcholine receptor antagonist</td>
<td>Interference with the release of reactive oxygen species</td>
</tr>
<tr>
<td>Strychnine-sensitive glycine receptors of wide dynamic range neurons</td>
<td>CNS inhibition via lidocaine mediated strychnine-sensitive glycine receptors of wide dynamic range neurons in the spinal cord → ↓ CMRO2, ↓ CBF → ↓ energy necessary for neuronal membrane stabilization</td>
</tr>
<tr>
<td>K+-Cl− cotransporter on γ-aminobutyric acid receptors</td>
<td>Blocks the K+-Cl− cotransporter on γ-aminobutyric acid receptors → progressive CNS symptoms</td>
</tr>
<tr>
<td>Protein kinase C</td>
<td>Inhibition of protein kinase C → ↓ phosphorylation of N-methyl-D-aspartate subunits</td>
</tr>
</tbody>
</table>

### Pharmacokinetics

The plasma concentration of lidocaine is dependent upon several factors including: dosage, absorption, distribution, elimination, patient age, patient hepatic function, patient cardiovascular status, and plasma protein binding. Lidocaine injection can be illustrated by a two-compartment model resulting in an initial rapid reduction of peak plasma concentrations (Cmax). The primary α-phase includes rapid uptake to the lungs and distribution to the highly vascular compartment. The subsequent β-phase results in a slow re-distribution to vascular poor regions. The vasodilatory effects of lidocaine contribute to its short duration of action and enhanced systemic absorption.
Central Nervous System Toxicity

Systemic lidocaine toxicity is a life-threatening emergency that begins with CNS symptoms, and if left untreated, may progress to cardiovascular toxicity and fatality. Lidocaine toxicity is dose dependent and largely influenced by Cmax. Cmax is directly influenced by the free fraction of lidocaine available. The free fraction of lidocaine is contingent upon plasma protein binding, total interstitial volume, and plasma volume. The extent of plasma protein binding is influenced by the serum pH and patient levels of alpha 1-glycoprotein and albumin. There is an indirect relationship between seizure threshold and the levels of serotonin (5-hydroxytryptophan) in the central nervous system. Thus, medications that elevated levels of 5-hydroxytryptophan may increase the risk for lidocaine CNS toxicity.

At a Cmax of 3-4 mcg/mL patients develop circumoral numbness, restlessness, tinnitus, tingling of the face, metallic taste in the mouth, auditory hallucinations, and visual hallucinations. At a Cmax of 5-10 mcg mL-1 tremors and seizure activity may ensue. Thus, intravenous lidocaine boluses should be titrated to a serum concentration less than 5-10 mcg mL-1, the seizure threshold for lidocaine. If toxicity remains untreated, it may progress to bradycardia, arrhythmia, hypotension, cardiac and respiratory arrest.

Management of Central Nervous System Toxicity

The initial management of lidocaine toxicity includes the following steps: call for back-up clinical support, stop lidocaine administration, intubation to secure the airway and administer a benzodiazepine. In a patient with no cardiovascular symptoms, propofol can also be administered at a 1 mg kg-1 IV. A 20% lipid emulsion may be administered at a 1.5 mL kg-1 followed by a continuous infusion of 0.25-0.5 mL kg-1 min-1. If the toxicity progresses to cardiovascular compromise, ACLS should be followed with the following modifications: use 10-100 mcg doses of epinephrine, utilize amiodarone for ventricular arrhythmias, avoid vasopressin, calcium channel blockers, beta-blockers, and consider cardiopulmonary bypass for patients that do not respond to treatment.

Discussion

The possible explanations for CNS toxicity in this case include: pharmaceutical interactions, cytochrome P450 3A4 mediated drug-drug interactions, concurrent beta-blockade, rate of administration and dosage.

1. Pharmaceutical Interactions: The patient’s home medications included tramadol and amloptiptan. Tramadol is a known noradrenaline and serotonin reuptake inhibitor. Almotriptan is a serotonin agonist. Tramadol and almotriptan enhance serotonergic transmission in the CNS, reducing the seizure threshold, consequently, increasing the risk for lidocaine-induced CNS toxicity. Increased serotonergic transmission in the CNS results in a reduction of the seizure threshold consequently increasing the risk for lidocaine related CNS toxicity.

2. Cytochrome P450 3A4 Mediated Drug-Drug Interactions: The patient’s home medication list includes the following competing substrates for CYP 450 3A4: oxycodone, cetirizine, and levonorgestrel. Moreover, benzodiazepines acts as a competing substrate. Simultaneous CYP450 3A4 metabolism increases Cmax. Medications that are simultaneously metabolized by CYP 450 3A4 are shown to increase risk of toxicity in patients administered lidocaine. This results an increase in Cmax. Likewise, simultaneous administration with known CYP450 3A4 inhibitors will result in an increase in Cmax.

3. Concurrent Beta-Blockade: Concurrent beta-blockade may also contribute to lidocaine CNS toxicity. Sympathetic suppression by small dose esmolol reduces blood flow to splanchic organs. The concomitant reduction of HR and decrease in hepatic blood flow and renal clearance, may result in an increased Cmax.

4. Bolus Dosing: A small study (n=8), showed that the administration a 1 mg kg-1 bolus dosing of intravenous lidocaine produces mild symptoms of CNS toxicity for 5 +/- 1.4 minutes duration including: circunoral numbness, metallic taste, dizziness, hallucinations, and slurred speech. It was identified, however, that the rapidity of exposure of lidocaine to the CNS had the greatest influence on the precipitation of CNS toxicity symptoms. The plasma concentration threshold where CNS toxicity occurs is more likely due to the rate of increase of the serum concentration than due to the total amount of drug injected.

5. Rate of Administration: The rate of lidocaine administration influences Cmax. A 2 mg kg-1 bolus of intravenous lidocaine theoretically results in a Cmax of 6.6-8.5 mcg/mL. The rate increase of Cmax likely has more influence on lidocaine toxicity than dosing. Rapid bolus administration of 2% lidocaine into the epidural space, has been shown to significantly (P<0.0005) increase Cmax when compared to a 20 mg min-1 infusion. There are no studies in the literature that review the effect on Cmax after intravenous bolus administration.

6. Genetic Polymorphism: In the absence of a normal therapeutic response, genetic polymorphism should be considered. There are 46 known single nucleotide polymorphisms of CYP450 3A4 affecting drug metabolism. Moreover, CYP450 3A4 enzyme phenotypic expression varies 30-fold between individuals.

Practice Recommendations

1. Caution with the co-administration of lidocaine with: SSRIs, SNRIs, serotonin agonists, antihistamines, beta-blockers, calcium channel blockers, macrolide antibiotics, antifungals, and benzodiazepines.
2. Caution with concurrent administration of intravenous lidocaine with known CYP 450 3A4 inhibitors and medications metabolized by CYP 450 3A4.
3. Consider dose reduction for intravenous lidocaine bolus administration.
4. Avoid the rapid bolus of intravenous lidocaine.
5. Genetic polymorphism should be considered when an abnormal therapeutic response is appreciated at standard doses.

Notes

The authors declare that no competing interests exist.

Ethical Declarations

Informed consent was obtained from the patient in this case study. All associated patient identifiers were removed for the purposes of this paper.

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